

The *in vivo* effect of *Rhodiola rosea* and *Rhodiola quadrifida* hydro-alcoholic extracts on chemokinetic activity of spleen lymphocytes in mice

EWA SKOPIŃSKA-RÓŻEWSKA¹, MAŁGORZATA BYCHAWSKA², BEATA BIAŁAS-CHROMIEC², EWA SOMMER¹

¹Department of Pathology, Biostructure Center, Warsaw Medical University; ²Department of Laboratory Diagnostics and Immunology, National Institute of Tuberculosis and Lung Research, Warsaw, Poland

Abstract

Rhodiola rosea (RR) and *Rhodiola quadrifida* (RQ) roots and rhizomes are traditional natural drugs used in Asia as adaptogens and antidepressants. The aim of this work was to study the *in vivo* effect of 50% hydro-alcoholic extracts of RR and RQ roots and rhizomes on the *ex vivo* chemokinetic activity of splenic lymphocytes in mice. Mice were fed for 7 days *Rhodiola* extracts in daily doses 40 or 200 µg. The chemokinetic activity of splenocytes was determined in 24-hour cell cultures in capillary tubes. Both extracts stimulated splenocytes mobility in lower dose, in higher dose (200 µg) only RQ extract was effective.

Key words: *Rhodiola rosea*, *Rhodiola quadrifida*, mice, splenocytes, chemokinesis.

(Centr Eur J Immunol 2009; 34 (1): 42-45)

Introduction

Rhodiola rosea (RR) and *Rhodiola quadrifida* (RQ) belong to the big family of adaptogenic herbs, comprising medicinal plants from various species – *Panax ginseng*, *Schizandra chinensis*, *Eleutherococcus senticosus* and many others. *Rhodiola rosea* is considered to be one of the most active adaptogenic drugs. Adaptogenic plants are used in traditional medicine to decrease depression, to enhance work performance and resistance to high-altitude illness, and to treat consequences of physical and psychological stress [1-7].

Besides of this stimulatory action on the nervous system, adaptogenic plants also have immunotropic activity [8-16]. Our earlier experimental studies documented immunomodulatory effects of extracts from *Panax ginseng*, *Eleutherococcus senticosus*, *Schizandra chinensis* and three *Rhodiola* species (*R. rosea*, *R. quadrifida* and *R. kirilowii*) on various parameters of immunological response *in vivo* and *in vitro* [17-30]. In the present paper we evaluate the

in vivo influence of *Rhodiola rosea* and *Rhodiola quadrifida* hydro-alcoholic extracts on chemokinetic activity of mice splenic lymphocytes *in vitro*.

Material and Methods

Rhodiola rosea (*Crassulaceae*) roots and rhizomes were cultivated, collected and identified in the Research Institute of Medicinal Plants (RIMP), Poznań, thanks to prof. Przemysław M. Mrozikiewicz and dr Waldemar Buchwald.

Rhizomes of *R. quadrifida* were collected in Altai mountain in Mongolia, thanks to dr H. Wiedenfeld. The Mongolian plant material was identified; voucher specimen was deposited at the herbarium of the Institute of Botany of Mongolian Academy of Science in Ulaanbatar. Samples for the study were obtained by Prof. Mirosława Furmanowa, head of the scientific project PBZ-KBN-092/PO5/2003. Sample extractions and their chemical analysis were performed by the scientists from RIMP (dr Alina Mścisz, dr Anna

Krajewska-Patan, dr Sebastian Mielcarek), and from Warsaw Medical University (prof. Mirosława Furmanowa, dr Małgorzata Hartwich, dr Marek Malinowski) as was described before [24, 25, 30]. Briefly: air-dried finely powdered roots and rhizomes were extracted two times with 50% ethanol (hydro-alcoholic extracts, RRA and RQA), at 40-45°C, evaporated to dryness and lyophilized. For the study of chemical composition of extracts HPLC methods have been used [31, 32]. Extracts were dissolved in 10% ethyl alcohol before administration to the animals.

The study was performed on 8-10-weeks old female inbred Balb/c mice, 20-22 g of body mass, delivered from the Polish Academy of Sciences breeding colony.

For all experiments animals were handled according to the Polish law on the protection of animals and NIH (National Institutes of Health) standards. All experiments were accepted by the local Ethical Committee.

Rhodiola extracts were administered to Balb/c mice *per os* in 7 daily doses of 40 or 200 µg (each group consisted of 8 mice). These doses corresponded to 20 or 100 mg given to 70 kg person (applying the coefficient equal 7 for adjusting differences between mouse and human in relation of the surface to body mass). Mice received drugs by Eppendorff pipette, in 40 µl of 10 % ethyl alcohol, for 7 days. Control mice (16 animals) were fed 40 µl of 10% ethyl alcohol. On the day 8th mice were bled in anaesthesia from retro-orbital plexus and sacrificed with Morbital. Splenocytes were isolated from spleens under sterile conditions by straining through stainless sieve and cotton gauze and centrifugation on Lymphoprep in order to remove erythrocytes.

Spleen cells chemokinesis (spontaneous migration) assay *in vitro* was performed according to the Sandberg method [33] in own modification [19, 23]. Briefly, isolated splenocytes were resuspended in Parker culture medium with 5% inactivated FCS, at the final concentration of 30×10⁶ cells/ml. Afterwards, siliconized capillary tubes were filled with cell suspension, sealed with plasticine, centrifuged (5 min 450 g) and fixed on the glass plates. Cells levels were marked. After 24 h incubation (37°C, 5% CO₂ humidified atmosphere) the distances of migration were measured in millimeters (mm) at a magnification of 6.5 × and presented as migration units (1 MU= 0.18 mm).

Stimulatory indices were calculated by dividing the results obtained for individual splenocytes cultures derived from *Rhodiola* fed animals by the mean of the results of accompanying control cultures.

Statistical evaluation of the results was done by 2-way analysis of variance (ANOVA) and Bonferroni post-test (GraphPad Prism software package).

Results

Performed analysis of variance revealed, that variation among column means is highly significantly greater than

expected by chance (Table 1). Bonferroni Multiple Comparison Test indicated differences between control group and groups of mice fed 0.04 mg daily dose of both extracts. Higher 0.2 mg dose was ineffective in the case of *Rhodiola rosea* (RRA) extract (Table 2 and Fig. 1).

Discussion

The aim of this study was to evaluate, for the first time, the *in vivo* effect of extracts obtained from the roots and rhizomes of *R. rosea* and *R. quadrifida* on chemokinetic activity of mice splenocytes in 24 h tissue cultures *in vitro*. The results obtained show higher *in vitro* migratory activity of splenocytes collected from mice fed lower dose of *Rhodiola* extracts. It may reflect the higher incidence in these cell suspensions of T lymphocytes, having better migratory properties than B cells.

Earlier experiments performed by us with some other plant adaptogens (extracts from *Eleutherococcus senticosus*, *Centella asiatica*, *Lithospermum canescens*) revealed stimulatory effect of lower doses and no effect or inhibitory influence of higher doses of these substances on mobility of splenocytes of treated mice [17, 19]. In the present study we observed similar effects. Moreover, our earlier studies on various other parameters of cellular specific and nonspecific immune response in mice, rats and pigs, revealed that *Rhodiola* extracts *in vivo* and *in vitro* enhanced these responses in lower doses and were ineffective or suppressed them in higher ones [24, 25, 27]. In experiments in mice, feeding cell donors 400 µg daily doses of *R. rosea* extracts

Table 1. Two-way ANOVA

Source of Variation	% of total variation	p value		
Interaction	5.90	p<0.0001		
Dose	53.36	p<0.0001		
Drug	8.61	p<0.0001		
Source of Variation	p value summary	Significant?		
Interaction	***	Yes		
Dose	***	Yes		
Drug	***	Yes		
Source of Variation	Difference	Sum-of-squares	Mean square	F
Interaction	2	0.3253	0.1627	15.60
Dose	2	2.943	1.471	141.2
Drug	1	0.4748	0.4748	45.54
Residual	170	1.772	0.01042	

Table 2. Bonferroni posttest

Control vs. 0.04				
Drug	Difference	t	p value	Summary
RRA	0.2300	7.748	p<0.001	***
RQA	0.4800	17.71	p<0.001	***
Control vs. 0.2 mg				
Drug	Difference	t	p value	Summary
RRA	0.01000	0.3369	p>0.05	ns
RQA	0.1300	4.379	p<0.001	***
0.04 vs. 0.2 mg				
Drug	Difference	t	p value	Summary
RRA	-0.2200	5.901	p<0.001	***
RQA	-0.3500	9.925	p<0.001	***

resulted in suppression of splenic lymphocytes angiogenic activity, what may suggest the presence of suppressor cells or production of suppressory cytokines. This dose corresponds to the doses recommended by producers of some dietary supplements which contain *Rhodiola rosea* (Anty-stress, Lentaya) [24]. In the case of *R. quadrifida* hydro-alcoholic extract, the best stimulation of lymphocytes angiogenic activity was obtained feeding mice 200 µg daily dose (what corresponds to 100 mg of human dose). Dose of 400 µg was not inhibitory, as in the case of *R. rosea*, but gave lower stimulatory effect [25]. It corresponds to the effects obtained by us for these two *Rhodiolas* in the present work.

Conclusion

In human, daily doses 20 mg, 50 mg (and in the case of *R. quadrifida* also 100 mg) of *Rhodiola* extracts would effectively stimulate lymphocyte-dependent immunity. Use of dietary supplements containing *R. rosea* in daily doses higher than 50 mg, might be dangerous because of the possibility of suppression of some lymphocyte activities. However, anti-bacterial activity of peritoneal macrophages increased in mice fed 400 µg of *Rhodiola rosea* extracts [in press], and metabolic activity of mice blood granulocytes fed *R. quadrifida* presented dose-dependent increase up to the 400 µg daily dose [28].

References

1. Monograph (2002): *Rhodiola rosea*. *Altern Med Rev* 7: 421-423.
2. Monograph (2006): *Eleutherococcus senticosus*. *Altern Med Rev* 11: 151-155.
3. Single Herb Monographs: Ginseng. <http://www.tufts.edu/med/ebcam/east-AsianMed/Ginseng/html>.

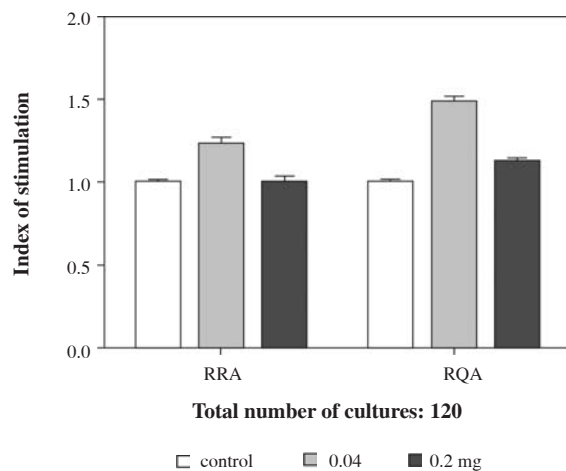


Fig. 1. The effect of feeding mice for 7 days *Rhodiola* hydro-alcoholic extracts on chemokinetic activity of splenic lymphocytes in 24-hours tissue culture (mean stimulation index \pm SEM)

4. Perfumi M, Mattioli L (2007): Adaptogenic and central nervous system effects of single doses of 3% rosavin and 1% salidroside *Rhodiola rosea* L. extract in mice. *Phytother Res* 21: 37-43.
5. Lee S, Kim DH, Jung JW et al. (2007): *Schizandra chinensis* and *Scutellaria balcalensis* counter stress behaviours in mice. *Phytother Res* 21: 1187-1192.
6. Panossian A, Wagner H (2005): Stimulating effect of adaptogens: an overview with particular reference to their efficacy following single dose administration. *Phytother Res* 19: 819-838.
7. Panossian A, Wikman G (2008): Pharmacology of *Schizandra chinensis* Ball: an overview of Russian research and uses in medicine. *J Ethnopharmacol* 118: 183-212.
8. Pooja BA, Khanum F (2009): Anti-inflammatory activity of *Rhodiola rosea* – “a second-generation adaptogen”. *Phytother Res* Jan 16 (Epub ahead of print).
9. Biondo PD, Robbins SJ, Walsh JD (2008): A randomized controlled crossover trial of the effect of ginseng consumption on the immune response to moderate exercise in healthy sedentary men. *Appl Physiol Nutr Metab* 33: 966-975.
10. Kormosh N, Laktionov K, Antoshechkina M (2006): Effect of a combination of extract from several plants on cell-mediated and humoral immunity of patients with advanced ovarian cancer. *Phytother Res* 20: 424-425.
11. Guo LY, Hung TM, Bae KH et al. (2008): Anti-inflammatory effects of schisandrin isolated from the fruit of *Schizandra chinensis* Ball. *Eur J Pharmacol* 59: 293-299.
12. Song X, Bao S, Wu L, Hu S (2009): Ginseng stem-leaf saponins (GSLs) and mineral oil act synergistically to enhance the immune responses to vaccination against foot-and-mouth disease in mice. *Vaccine* 27: 51-55.
13. Lim DS, Bae KG, Jung IS et al. (2002): Anti-septicaemic effect of polysaccharide from *Panax ginseng* by macrophage activation. *J Infect* 45: 32-38.
14. Wang H, Actor JK, Indrigo J et al. (2003): Asian and Siberian ginseng as a potential modulator of immune function: an in vitro cytokine study using mouse macrophages. *Clin Chim Acta* 327: 123-128.

15. Liou CJ, Huang WC, Tseng J (2006): Short-term oral administration of ginseng extract induces type-1 cytokine production. *Immunopharmacol Immunotoxicol* 28: 227-240.
16. Chen TS, Liou SY, Chang YL (2008): Antioxidant evaluation of three adaptogen extracts. *Am J Chin Med* 36: 1209-1217.
17. Pietrosiuk A, Skopińska-Różewska E, Furmanowa M (2004): Immunomodulatory effect of shikonin derivatives isolated from *Lithospermum canescens* on cellular and humoral immunity in Balb/c mice. *Pharmazie* 59: 640-642.
18. Furmanowa M, Skopińska-Różewska E, Rogala E, Hartwich M (1998): *Rhodiola rosea* in vitro culture-phytochemical analysis and antioxidant action. *Acta Soc Botanic Pol* 67: 69-73.
19. Rogala E, Skopińska-Różewska E, Sawicka T et al. (2003): The influence of *Eleutherococcus senticosus* on cellular and humoral immunological response of mice. *Pol J Vet Sci* 6 (Suppl): 37-39.
20. Skopińska-Różewska E, Nartowska J, Augustynowicz J et al. *Żeńszeń w świetle współczesnych badań naukowych*. In: *Immunomodulacja – nowe możliwości w ochronie zdrowia*. Eds. Siwicki AK, Skopińska-Różewska E, Świdorski F. SPW EDYCJA, Olsztyn 2004; pp. 21-27.
21. Siwicki AK, Skopińska-Różewska E, Nartowska J et al. (2004): Effect of Immunostim plus – a standardized fixed combination of *Schizandra chinensis* with *Eleutherococcus senticosus* extracts on granulocyte activity and tumor angiogenesis in mice. *Bull Vet Inst Pulawy* 48: 489-492.
22. Rogala E, Sommer E, Radońska-Leśniewska D et al. (2004): Immunomodulatory effects of *Panax ginseng* preparations on the mouse. *Herba Polonica* 50: 38-44.
23. Skopińska-Różewska E, Siwicki AK, Wójcik R et al. (2006): Immunostimulatory effect of Immunostim-plus – a standardized fixed combination of *Schizandra chinensis* with *Eleutherococcus senticosus* extracts on lymphocyte – dependent cellular immunity in mice. *Bull Vet Inst Pulawy* 50: 461-465.
24. Siwicki AK, Skopińska-Różewska E, Hartwich M et al. (2007): The influence of *Rhodiola rosea* extracts on non-specific and specific cellular immunity in pigs, rats and mice. *Centr Eur J Immunol* 32: 84-91.
25. Skopińska-Różewska E, Wójcik R, Siwicki AK (2008): The effect of *Rhodiola quadrifida* extracts on cellular immunity in mice and rats. *Pol J Vet Sci* 11: 105-111.
26. Skopińska-Różewska E, Malinowski M, Wasutyński A et al. (2008): The influence of *Rhodiola quadrifida* 50% hydro-alcoholic extract and salidroside on tumor-induced angiogenesis in mice. *Pol J Vet Sci* 11: 97-104.
27. Wójcik R, Siwicki AK, Skopińska-Różewska E et al. (2008): The in vitro influence of *Rhodiola quadrifida* extracts on non-specific cellular immunity in pigs. *Centr Eur J Immunol* 33: 193-196.
28. Skopińska-Różewska E, Bychawska M, Sommer E, Siwicki AK (2008): The in vivo effect of *Rhodiola quadrifida* extracts on the metabolic activity of blood granulocytes in mice. *Centr Eur J Immunol* 33: 179-181.
29. Skopińska-Różewska E, Wasutyński A, Sommer E et al. (2008): The influence of *R. rosea*, *R. kirilowii* and *R. quadrifida* extracts on cutaneous angiogenesis induced in mice after grafting of human kidney cancer tissue. *Centr Eur J Immunol* 33: 185-189.
30. Skopińska-Różewska E, Hartwich M, Siwicki AK et al. (2008): The influence of *Rhodiola rosea* extracts and rosavin on cutaneous angiogenesis induced in mice after grafting of syngeneic tumor cells. *Centr Eur J Immunol* 33: 102-107.
31. Mielcarek S, Mścisz A, Buchwald W et al. (2005): Phytochemical investigation of *Rhodiola* sp. Root extracts. *Herba Polonica* 51 (Suppl 1): 159.
32. Wiedenfeld H, Dumaa M, Malinowski M, Furmanowa M (2007): Phytochemical and analytical studies of extracts from *Rhodiola rosea* and *Rhodiola quadrifida*. *Pharmazie* 62: 308-311.
33. Sandberg G (1976): The sealed capillary migration technique and thymocyte migration in vitro. *J Immunol Meth* 12: 365-368.