

Molecular cloning and sequencing analysis of the interferon receptor (IFNAR-1) from *Columba livia*

CHAO LI, WEI SHAN CHANG

Shandong Agriculture University, Shandong, PR China

Abstract

Objective: Partial sequence cloning of interferon receptor (IFNAR-1) of *Columba livia*.

Material and methods: In order to obtain a certain length (630 bp) of gene, a pair of primers was designed according to the conserved nucleotide sequence of *Gallus* (EU477527.1) and *Taeniopygia guttata* (XM_002189232.1) IFNAR-1 gene fragment that was published by GenBank. Special primers were designed by the Race method to amplify the 3' terminal cDNA.

Results: The *Columba livia* IFNAR-1 displayed 88.5%, 80.5% and 73.8% nucleotide identity to *Falco peregrinus*, *Gallus* and *Taeniopygia guttata*, respectively. Phylogenetic analysis of the IFNAR1 gene showed that the relationship of *Columba livia*, *Falco peregrinus* and chicken had high homology.

Conclusions: We successfully obtained a *Columba livia* IFNAR-1 gene partial sequence. Analysis of the genetic tree showed that the relationship of *Columba livia* and *Falco peregrinus* IFNAR-1 had high homology. This result can be used as reference for further research and practical application.

Key words: *Columba livia*, IFNAR-1, gene cloning, gene sequence analysis.

(Centr Eur J Immunol 2014; 39 (2): 159-164)

Introduction

Interferon was identified by Isaacs and Lindenmann during their seminal studies on virus interference 50 years ago. They reported that influenza virus-infected chick cells produced a secreted factor that transferred a virus-resistant state to previously uninfected cells. Importantly, the resistance was displayed against both the homologous inducing virus and heterologous viruses. The factor was designated interferon (IFN) because of its ability to interfere with virus growth [1]. Since then IFN has been implicated in a wide variety of biological functions, including anti-proliferative effects, induction of human leukocyte antigen (HLA), increase in natural killer (NK) activity and others [2, 3].

Interferon proteins are historically characterised as type I (α , β) or viral IFNs and type II (γ) or immune IFN [4]. Type I IFN induces antiviral, anti-proliferation, and immunomodulatory effects through interaction with specific receptor complexes on the surface of target cells, which consists of two chains: The high-affinity ligand-subunit (IFN- α , β receptor α -chain, IFNAR-1) and the unknown function subunit (IFN- α , β receptor β -chain, IFNAR-2) [5, 6]. Similarly to other IFN receptors, IFNAR-1 is a single transmembrane protein of the class II

cytokine receptor family (CRF2), defined by structural similarities in the extracellular domain. Interferon receptor-mediated signalling leads to the activation of latent cytoplasmic factors, the signal transducers and activators of transcription (STAT) family of proteins, which are activated by a process that involves members of the Janus tyrosine kinase (JAK) family [7-9].

The objective of this study was to clone a sequence of IFNAR-1 gene and provide a basic understanding of the gene mutations between *Columba livia* and animals closely related to it through phylogenetic analysis. Physiological and biochemical property studies of the obtained IFNAR-1 gene and protein were conducted to provide valuable reference for further research on the gene.

Material and methods

Sample collection and tissue preparation

Spleen samples were collected from *Columba livia*. The *Columba livia* was brought as a live, mature *Columba livia* from a common farm and kept under inspection for 5 days to be sure that it was free from any clinical infection, then the sample was harvested and stored at -20°C .

Table 1. Conserved sequence amplification PCR and 3'RACE products using forward and reverse primer sequences

Primer name	Primer sequences
A	5'-GCTCAGATTGGTCCCCCTGA-3' 5'-CAGGAGGGCCAATTTTCATTTGTT-3'
3'RACE 1	5'-GCACCAGACACTACCTATT-3' 5'-CTAGCGCTCCTGACTCAGCGGTTTTTTTTTTTTTTTTTTTT-3'
3'RACE 2	5'-ACTCCACAGAGTGGCTCAGTGT-3' 5'-CTAGCGCTCCTGACTCAGCGG-3'

Total RNA isolation extraction and synthesis of cDNA

Total RNA samples were extracted from spleens using Trizol (TransGen), and the cDNA was obtained using the PrimScript RT reagent Kit (TaKaRa) according to the manufacture's protocol.

RT-PCR, 3'RACE and sequencing

A pair of homologous primers (Table 1A) was designed with DNASTAR 5.0 software in the conserved region of Gallus (EU477527.1) and Taeniopygia guttata (XM_002189232.1). All primers used in this study are

listed in Table 1. With the primers, a cDNA fragment was amplified by RT-PCR using the first strand cDNAs as templates. The PCR was performed at 94°C for 5 minutes, followed by 30 cycles at 94°C for 45 seconds, 58°C for 40 seconds and 72°C for 1 minute, with a final extension at 72°C for 10 minutes, and then stopped at 4°C. Polymerase chain reaction products were detected by electrophoresing 5 µl aliquots through 1% agarose in 1 × TAE and purified by Agarose Gel DNA Extraction Kit (Shanghai Sangon Biotech Co., Ltd.). The products were cloned by into pEASY-T1 vector (TransGen) and sent to Shanghai Sangon Biotech Co., Ltd. for sequencing. Two pairs of specific primers (Table 1: 3'RACE 1 and 2) were designed

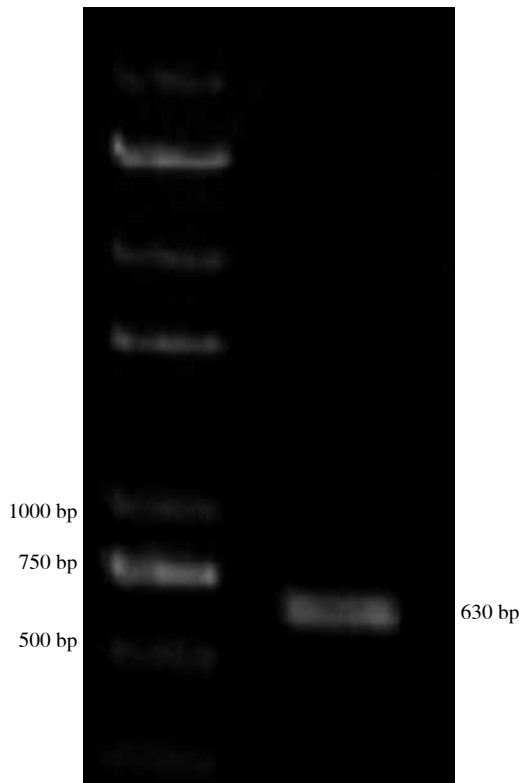


Fig. 1. RT-PCR amplification result of the conserved sequence

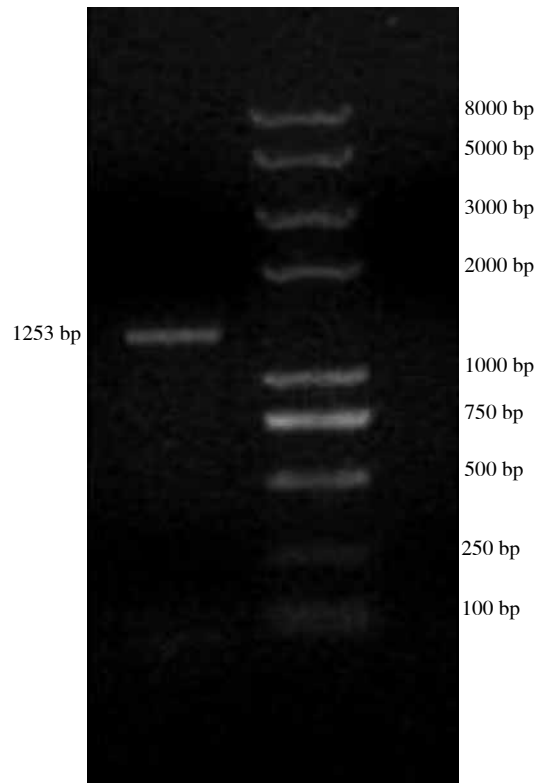


Fig. 2. Results of 3'RACE amplification

according to the obtained sequences. In the 3'RACE and the second pair for the second round PCR products were analysed by electrophoresis in 1% agarose then purified and cloned into pEASY-T1 vector, detected and sequenced as mentioned above.

IFNAR1 nucleotide sequence homology comparison and phylogenetic analysis

Sequencing results of *Columba livia* IFNAR-1 and the nucleotide sequence of *Gallus* (EU477527.1), *Falco peregrinus* (XM_005234387.1), *Melopsittacus undulatus* (XM_005151790.1), *Anas platyrhynchos* (XM_005013847.1), *Ficedula albicollis* (XM_005037095.1), *Taeniopygia guttata* (XM_002189232.1), *Papio Anubis* (NM_001168783.1), *Macaca mulatta* (NM_001266513.2), *Homo sapiens* (XM_005260964.1), *Geospiza fortis* (XM_005427794.1) that registered in GenBank were sub-typed and put up for homologous analysis.

IFNAR1 amino acid analysis

Phylogenetic analysis was made using DNASTAR software. Sequence analysis of the predicted *Columba livia* IFNAR-1 protein translated from the nucleotide sequence of the *Columba livia* IFNAR1 fragment was performed using NCBI and ExPaSy software. Signal peptides were predicted using the Signal P 3.0 server. The domain structure of the Caprine protein was analysed on the SMART server. TMHMM Server v2.0 was used to predict transmembrane regions of IFNAR-1 protein.

Results

Molecular cloning and analysis of *Columba livia* IFNAR-1

A cDNA fragment of *Columba livia* IFNAR-1 of 630 bp (Fig. 1) was obtained from single RT-PCR that was

	1	2	3	4	5	6	7	8	9	10	11		
1	█	88.5	73.8	80.5	74.0	55.8	55.9	86.0	56.2	73.8	80.3	1	Columba livia.seq Falco peregrinus.seq Ficedula albicollis.seq gallus.seq Geospiza fortis.seq Homo sapiens.seq Macaca mulatta.seq Melopsittacus undulatus.seq Papio anubis.seq Taeniopygia guttata.seq Anas platyrhynchos.seq
2	12.6	█	72.1	79.9	71.5	56.4	55.9	86.3	57.1	70.3	81.6	2	
3	32.8	35.5	█	68.3	89.9	46.3	46.2	71.3	49.3	88.2	66.7	3	
4	22.8	23.7	42.0	█	68.6	54.5	54.3	77.9	55.5	67.2	79.2	4	
5	32.3	36.6	10.9	41.6	█	48.9	49.1	71.5	49.4	90.5	69.6	5	
6	68.3	66.9	99.6	71.9	90.2	█	90.6	55.4	95.4	46.6	51.4	6	
7	67.9	68.4	100.0	72.4	89.6	10.1	█	54.7	98.9	47.9	52.2	7	
8	15.8	15.3	36.9	26.5	36.5	69.5	71.6	█	57.1	68.7	81.3	8	
9	67.2	65.4	88.5	69.0	88.2	4.7	1.1	65.0	█	49.5	57.0	9	
10	32.7	38.4	12.8	43.7	10.2	98.1	92.5	41.2	87.4	█	68.4	10	
11	23.1	21.3	44.8	24.7	39.6	79.9	77.6	21.7	65.4	41.6	█	11	
	1	2	3	4	5	6	7	8	9	10	11		

Fig. 3. Homology comparisons of nucleotide sequences among *Gallus*, *Columba livia*, *Falco peregrinus* and other animals IFNAR-1 from GenBank

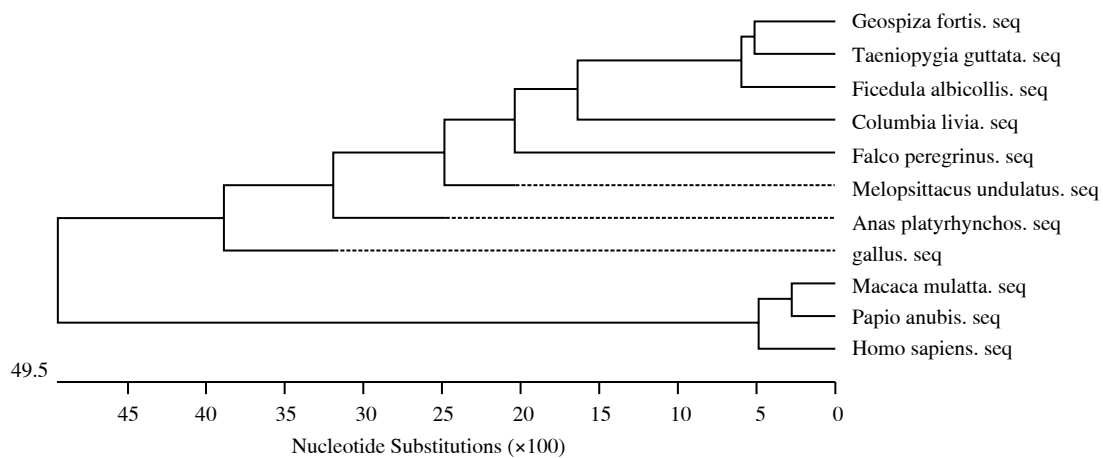


Fig. 4. The evolutionary tree of *Columba livia* compared with other animals from GenBank

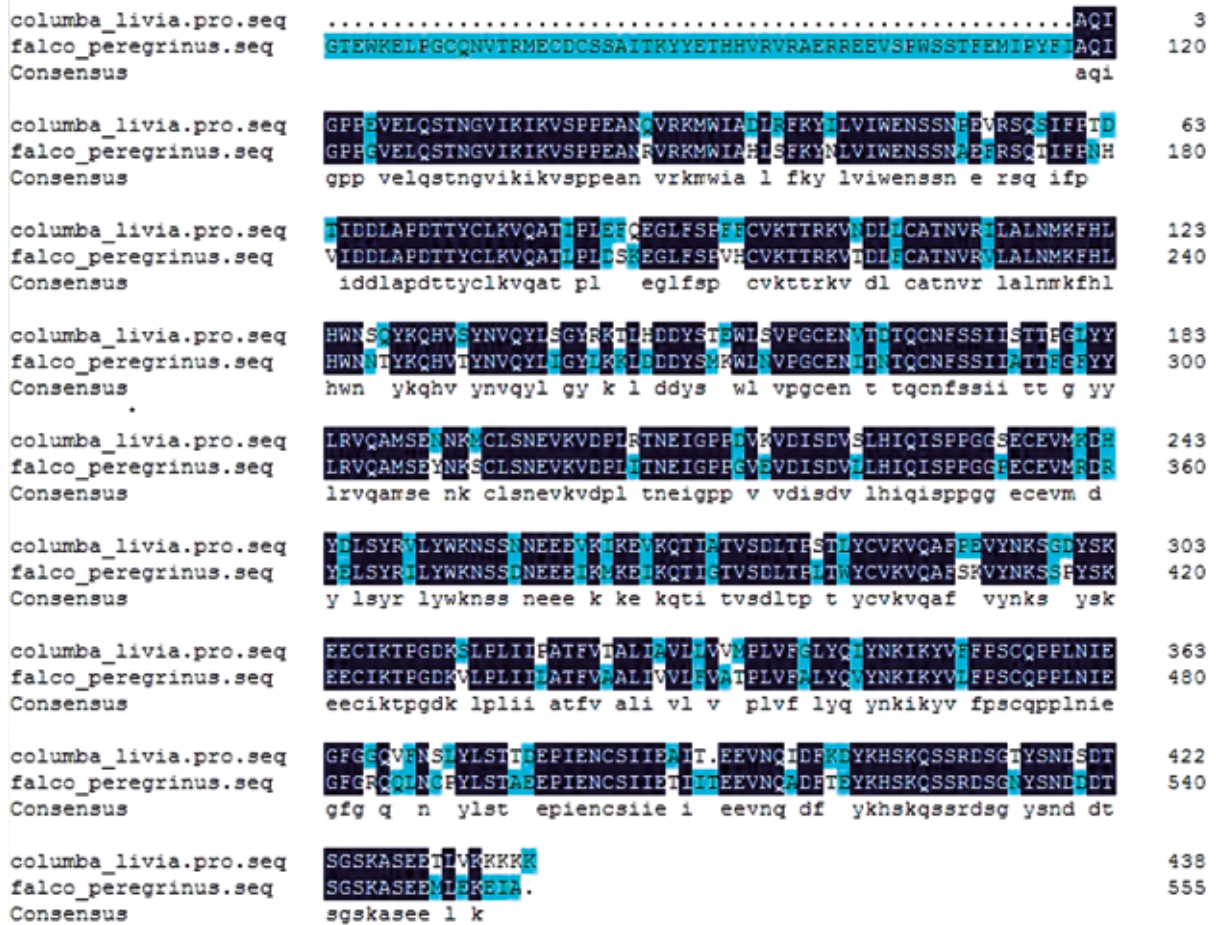


Fig. 5. Comparison of the predicted amino acid sequences of *Columba livia* and *Falco peregrinus* IFNAR-1. The blue shade reveals the conserved amino acid residues and the green shade reveals the non-conserved amino acid residues

	1	2	3	4	5	6	7	8	9	10	11	
1	69.7	72.7	57.6	69.3	61.6	32.5	34.7	71.2	39.8	61.7	1	Anas platyrhynchos.pro
2	38.7	80.4	67.1	68.0	68.5	36.8	36.4	77.3	37.1	68.8	2	columba livia.pro
3	33.9	22.8	66.7	68.5	66.5	38.8	39.2	77.8	40.4	65.7	3	falco peregrinus.pro
4	61.5	43.1	43.9	59.5	85.7	30.4	31.8	64.7	35.0	80.4	4	Ficedula albicollis.pro
5	39.5	41.7	40.8	57.7	60.0	35.7	36.4	67.0	37.3	58.8	5	gallus.pro
6	53.3	40.8	44.3	15.9	56.6	32.9	34.3	65.1	34.2	83.9	6	Geospiza fortis.pro
7	145.5	124.6	115.9	157.5	129.4	143.2	83.4	36.7	91.4	31.2	7	Homo sapiens.pro
8	134.1	126.0	114.6	149.0	126.4	135.8	18.9	36.9	97.3	32.5	8	Macaca mulatta.pro
9	36.2	27.0	26.3	47.5	43.3	46.7	124.8	124.2	38.4	62.0	9	Melopsittacus undulatus.pro
10	112.3	123.2	110.1	132.7	122.1	136.3	9.1	2.7	117.8	34.6	10	Papio anubis.pro
11	53.1	40.3	45.7	22.8	59.0	18.2	152.8	145.5	52.6	134.3	11	Taeniopygia guttata.pro
	1	2	3	4	5	6	7	8	9	10	11	

Fig. 6. Homology comparisons of amino acid sequences among *Gallus*, *Columba livia*, *Falco peregrinus* and other animals IFNAR-1 from GenBank

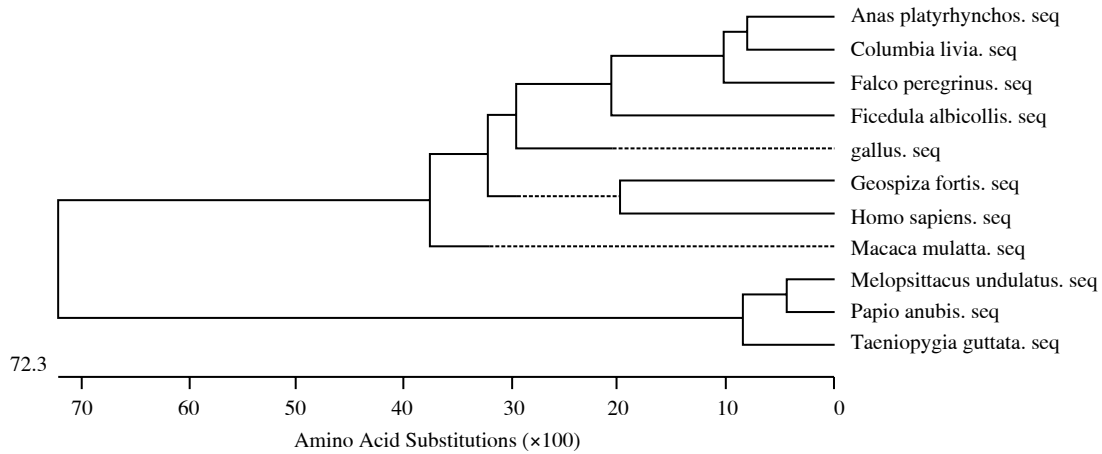


Fig. 7. The evolutionary tree of *Columba livia* compared with other animals from GenBank

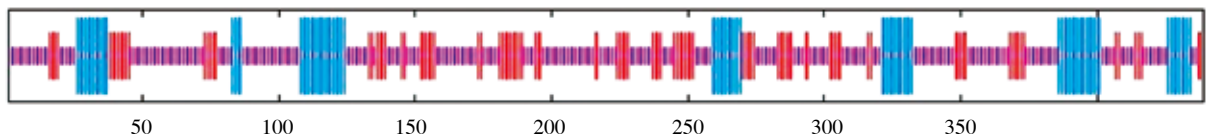


Fig. 8. The predicted secondary structure of *Columba livia* IFNAR-1 amino acid sequence. The blue line represents α -helix, the red line represents extended strand and the purple line represents random coil

identified from the spleen cDNA library of *Columba livia*, which are homologous to *Gallus* and *Taeniopygia guttata*. Based on this sequence, the 3' terminal cDNA was cloned using 3'RACE PCR. After splicing, a 1253 bp fragment (Fig. 2) of *Columba livia* IFNAR-1 was obtained.

IFNAR-1 sequence analysis

The *Columba livia* IFNAR-1 displayed 88.5%, 80.5% and 73.8% nucleotide identity to *Falco peregrinus*, *Gallus* and *Taeniopygia guttata*, respectively (Figs. 3 and 4). Analysis of the phylogenetic tree showed that the relationship of *Columba livia* and *Falco peregrinus* IFNAR-1 had high homology.

IFNAR1 amino acid analysis

The deduced amino acid (aa) sequences of *Columba livia* IFNAR-1 had an estimated isoelectric point and Mr of 5.55 and 50.0 KD, respectively. Multiple sequence alignment was carried out using the software package MegAlign (Lasergene 7.0, DNASTar Inc., Madison, WI). The *Columba livia* IFNAR1 displayed 80.4% and 68% aa identity to *Falco peregrinus* and *Gallus*, respectively. The inferred aa sequences of the IFNAR-1 of *Columba livia* and *Falco peregrinus* were aligned (Fig. 5). There were few amino acid mutations between *Columba livia* and *Falco peregrinus*; therefore, they had high homology. Figures

6 and 7 show the distance-based neighbour-joining tree displaying the phylogenetic relationships among *Columba livia* and other animals based on the *IFNAR1* gene. *Columba livia* has a closer relationship with birds such as *Falco peregrinus* and *Gallus*, and it has a more distant relationship with mammals. The secondary structure of the sequence was predicted to be mainly random coil, separated by extended strand and α -helix (Fig. 8). Transmembrane regions were predicted to be outside between the first and

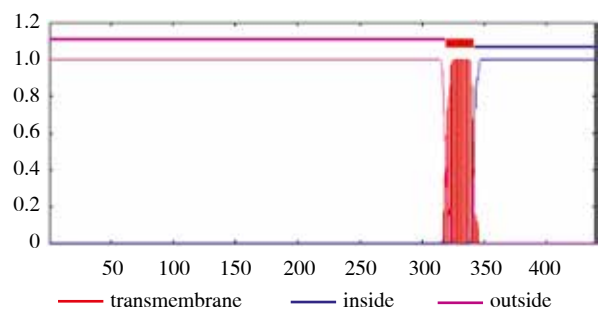


Fig. 9. Analysis of transmembrane regions of *Columba livia* IFNAR-1. The blue line represents inside, the red line represents transmembrane and the purple line represents outside

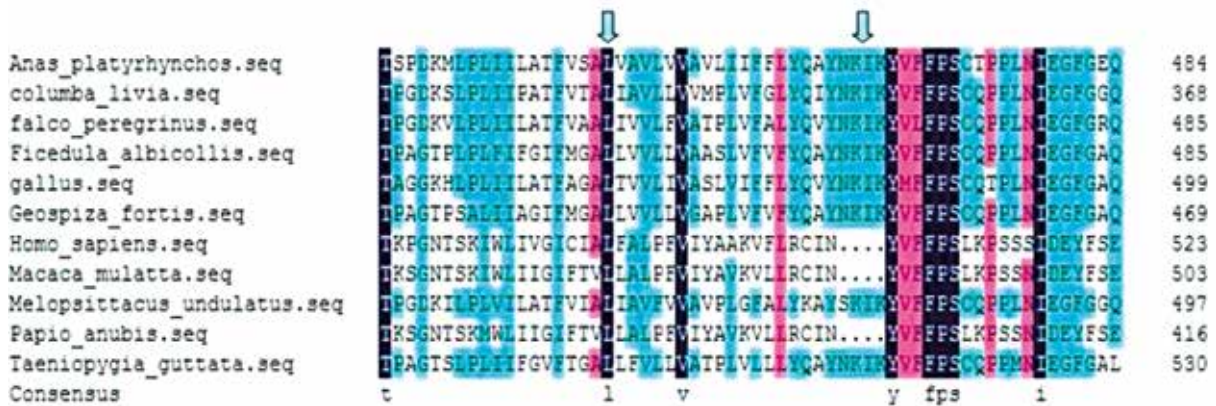


Fig. 10. Comparison of the amino acid sequences of several avian and mammalian species. The region between the two arrows represents the transmembrane region

318th amino acid, and transmembrane between 319th and 341th, and inside between the 342th and 439th (Fig. 9).

Discussion

According to the results, we can determine that *Columba livia* IFNAR-1 had high homology with avian, while its identity to mammalian was low. The highest homology was with *Falco peregrinus* (88.5%), the second was with *Melopsittacus undulates* (86%) and the third was with *Gallus* (80.5%).

According to the analysis of the amino acid sequences of the species by TMHMM Server v2.0, the transmembrane regions are similar. They all include approximately 20 amino acids that are all hydrophobic amino acids, such as Ala, Leu (Fig. 10).

The meat of *Columba livia*, known as “Dragon-meat”, is delicious in taste and well received in China, especially as squab. The number of *Columba livia* being raised is about 50 million and it is worldwide in distribution. *Columba livia* is an important part of China’s special economic animal production. But the occurrence of avian influenza came to be a serious threat to the production of *Columba livia*. It is necessary for *Columba livia* breeding to establish an immune mechanism, to find a new and an efficient security immune route. By contrast, the research of interferon receptor lags behind other areas of interferon research. In this study, we successfully obtained a *Columba livia* IFNAR-1 partial sequence and analysed the gene sequence and the amino acid sequences. The result can be used as reference for further research and practical application.

The authors declare no conflict of interest.

References

1. Isaacs A, Lindenmann J (1957): Virus interference. I. The interferon. Proc R Soc Lond B Biol Sci 147: 258-267.
2. Altmann SM, Mellon MT, Distel DL, Kim CH (2003): Molecular and functional analysis of an interferon gene from the zebrafish, *Danio rerio*. J Virol 77: 1992-2002.
3. Sen GC (2001): Viruses and interferons. Annu Rev Microbiol 55: 255-281.
4. Shtrichman R, Samuel CE (2001): The role of gamma interferon in antimicrobial immunity. Curr Opin Microbiol 4: 251-259.
5. Colamonici OR, D’Alessandro F, Diaz MO, et al. (1990): Characterization of three monoclonal antibodies that recognize the interferon α receptor. Proc Natl Acad Sci U S A 87: 7230-7234.
6. Colamonici OR, Domanski P (1993): Identification of a new sub unit of the type I IFN receptor localized to human chromosome 21. J Biol Chem 268: 10895-10899.
7. Samuel CE (2007): Interferons, interferon receptors, signal transducer and transcriptional activators, and interferon regulatory factors. J Biol Chem 282: 20045-20046.
8. Taniguchi T, Takaoka A (2002): The interferon- α/β system in antiviral responses: a multimodal machinery of gene regulation by the IRF family of transcription factors. Curr Opin Immunol 14: 111-116.
9. Müller M, Briscoe J, Laxton C, et al. (1993): The protein tyrosine kinase AK1 complements defects in the interferon- α/β and - γ signal transduction. Nature 366: 129-135.