Investigations of cellular immunity in juvenile idiopathic arthritis

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Abstract

The following was emphasised in an informative, educational issued on the American College of Rheumatology website in April 2017: "About one child in every 1000 develops some type of chronic arthritis. These disorders can affect children at any age, although rarely in the first six months of life. It is estimated that around 300,000 children in the United States have been diagnosed with the condition". Therefore, knowledge of immunological investigations in patients with juvenile idiopathic arthritis is important for finding new treatment pathways. Our aim was to assess the immunological investigations and immune system implications in juvenile idiopathic arthritis. We will discuss: a) the specifically targeted proteins – the citrullinated peptide antibodies; b) non-specifically targeted proteins – heat-shock proteins (anti-HSP60, -65, and -70 antibodies), CLEC16A, inflammasomes, and phagocyte-derived \$100; c) interleukins – IL-1, IL-6, IL-10, IL-17, and IL-18; d) innate immunity – macrophage activation syndrome, natural killer cells, complement activity, and immune complexes; and e) therapeutic targets – monoclonal antibodies, JAK inhibitors, and intravenous immune globulin.

Key words: juvenile idiopathic arthritis, immunopathology, immunological targets.

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Introduction

The pathological mechanism of autoinflammatory diseases includes the activation of the innate immune system associated with the lack of autoantibody synthesis [1]. The connections between autoinflammation and autoimmunity favour medical means of treating autoinflammatory diseases [2, 3]. Juvenile idiopathic arthritis (JIA) represents the "classic" autoinflammatory disorder [4]. Therefore, an immunological approach of the pathological mechanism in JIA is necessary.

Specifically targeted proteins

Citrullinated peptides (CCP) were specifically targeted in JIA. Hromadnikova *et al.* analysed the presence of IgG anti-CCP antibodies and IgG anti-keratin antibodies in sera of JIA patients at more than one year after diagnosis. Thus, a rare occurrence of anti-CCP was found [5]. Later, Wu *et al.* concluded that the clinical specificity of anti-CCP3 (third generation) was lower than that of the anti-CCP2 assay in JIA diagnosis, due to the cross-reaction in patients with telangiectasia syndrome [6]. Habib *et al.* found that

anti-CCP antibodies are correlated with joint erosions in JIA patients (p = 0.004) [7]. Anti-CCP antibodies, as non-systemically reacting antibodies, were intensively studied in JIA [8]. Wang *et al.* found a high specificity of 99.0% (95% CI: 98.0-100.0%) of anti-CCP antibody for the diagnosis of JIA [9]. Therefore, CCP are specifically targeted by autoantibodies [10].

Non-specifically targeted proteins

Heat stock proteins

The relationship between bacterial heat shock proteins (HSP) and autoimmunity was first disclosed in the *Mycobacterium bovis* (MB)-induced model of adjuvant arthritis [11]. Zlacka *et al.* studied the frequency of anti-HSP60, -65, and -70 antibodies in the sera of JIA patients. They found that the number of JIA patients (16/209, 7.6%) with elevated anti-HSP65 antibodies was equal to that of the healthy controls (4/50, 8%) [12]. Nguyen *et al.* found that the sera of JIA patients reacted with individual MB-HSP65 fragments P1-163 and P290-534. The levels of these fragments were increased in JIA patients compared to

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the healthy controls [13]. The immune response against self hsp65 in autoimmune arthritis was found to be protective rather than pathogenic [14]. On the other hand, the inhibition of regulatory T (Treg) cells, such as natural Foxp3(+) Treg and self HSP-induced Treg cells, along with a decreased amount of anti-inflammatory cytokine IL-10, results in the loss of immune tolerance [15]. These insights into HSP65 immunity would not only advance our understanding of the disease process in JIA, but also lead to the development of novel therapeutic approaches for autoimmune arthritis [16].

CLEC16A

CLEC16A was not associated with the susceptibility to anti-CCP-positive rheumatoid arthritis (RA) [17]. However, we found a functional link between human CLEC16A variation and the risk of autoimmunity [18].

Inflammasomes

Inflammasomes are multi-protein complexes composed of a NOD-like receptor (NLR)/an AIM-like receptor (ALR), and the adapter molecule apoptosis-associated speck-like protein, which contains a CARD (ASC), and caspase-1. The NOD-like receptor family, pyrin domain containing 1 (NLRP1) haplotypes, contributes to the susceptibility of developing vitiligo. There are other single nucleotide polymorphisms (SNPs) that alter the susceptibility and severity of JIA [19]. Recent studies conclude that inappropriate recognition of cytoplasmic self-DNA by AIM2 contributes to the development of psoriasis, dermatitis, arthritis, and other autoimmune and inflammatory diseases [20].

Phagocyte-derived S100

Phagocyte-derived S100 proteins are valid tools in the diagnosis of autoinflammatory diseases. In addition, these proteins may help to gain a better understanding of the pathophysiology of autoinflammatory disorders such as systemic juvenile idiopathic arthritis (sJIA) [21]. sJIA can be distinguished from other forms of JIA that usually manifest as a milder phenotype. S100 protein complexes enhance the pro-inflammatory phenotype [22].

Interleukins

The efficacy of the interleukin (IL)-1 and IL-6 as inhibiting agents in sJIA was debated [23]. IL-6 plays a significant role in many rheumatological diseases and has been described as both a pro- and anti-inflammatory cytokine [24]. IL-6 is thus crucially involved in the regulation of immune responses, haematopoiesis, and inflammation. When infections and tissue injuries occur, IL-6 is promptly synthesised in order to perform a protective role in the host's defence against stress or trauma. A humanised anti-IL-6

receptor monoclonal antibody has been proven to be outstandingly efficacious against JIA [25]. IL-1 and IL-6 play a major role in the pathogenesis of sJIA, thus the treatment using IL-1 and IL-6 inhibitors has been shown to be highly effective [26]. Tocilizumab (TCZ), a humanised anti-IL-6 receptor antibody, was developed [27]. IL-6 blockers were used in systemic onset JIA. TCZ, the first humanised anti-human IL-6 receptor antibody, inhibits the activity of IL-6 [28]. IL-10 gene polymorphism was associated with susceptibility to JIA. Harsini et al. observed no differences in the frequency of alleles, genotypes, and haplotypes of the IL-10 gene between groups of patients and controls [29]. However, Fathy et al. found a significant positive association between the IL-10 -1082 AA gene variant and susceptibility to polyarticular juvenile idiopathic arthritis (pJIA) [30]. A statistically significant increase in TNF-α, IFN-γ, IL-10, and IL-17 levels was found in children with JIA [31]. IL-17A was also prevalent in sera from patients with active sJIA, but the pathophysiological role of IL-17 is still unknown [32]. Another interleukin studied in sJIA was IL-18 [33]. Patients with sJIA shared a similar cytokine profile pattern characterised by a significant increase in IL-18 [33]. Myeloid-related protein (MRP) 8, MRP14, S100A12, and IL-18 are already being used as markers for active sJIA. Furthermore, in the case of non-sJIA subtypes, different markers such as HLA-B27, antinuclear-antibodies, RF, erythrocyte sedimentation rate, and C-reactive protein represent a resource for disease classification, prognosis, and activity. Ongoing studies are assessing the clinical role of MRP8, MRP14, and S100A12 [34]. Shimizu et al. observed that IL-18 might play a key role in the pathogenesis of macrophage activation syndrome (MAS). A serum IL-18 level greater than 47750 pg/ml might be a useful marker for predicting MAS development [35].

Innate immunity

MAS and natural killer cells

MAS, known as secondary haemophagocytic lymphohistiocytosis, is a complication of many rheumatic diseases, most commonly sJIA [36] - 10% of children with sJIA develop MAS. However, MAS occurs subclinically in another 30-40% [37]. Natural killer (NK) cells are activated early during inflammatory events in order to help shape the ensuing adaptive immune response. De Matos et al. concluded that CD94/NKG2A represents a key regulator in the synovial NK-cell cytokine synthesis, resulting in an activated phenotype of synovial NK-cell cytokine [38]. Defects of an unknown cause in the NK cell's cytotoxic capacity are presumed to underlie the pathogenesis of MAS, and they have been detected in sJIA patients [39]. Patients with JIA-ERA (enthesitis related arthritis) have an increased frequency of NK cells (12.89% ±5.65%) compared to healthy controls (9.34% $\pm 3.06\%$; p = 0.019) and diseased controls (8.81% $\pm 4.73\%$; p = 0.01) [40]. The pathogenesis of MAS may be related to the decrease in NK cell activity. The most consistent immunological abnormality reported in these patients is the impairment of cytotoxic functions. However, the detailed mechanism of this condition, including a clear role of NK cell dysfunction, is still being studied [41].

Complement activity and immune complexes

Complement activity was related to thymocytotoxic activity. The presence of thymocytotoxic activity was tested in synovial fluid obtained from JIA patients [42]. The rheumatoid factor cross-reactive idiotype (RF-CRI) was expressed in high concentrations in the sera of some patients with JIA. However, Bonagura et al. found an increased expression of RF-CRI in systemic lupus erythematosus (SLE) patients, which correlated inversely with C3 serum levels (r = 0.3925, p < 0.05) [43]. Jarvis *et al*. used two methods of sequential column chromatography to purify immune complexes from the synovial fluids of children with JIA. They demonstrated that high molecular weight complexes, containing IgM-RF, have not bound C4 in vivo, but activate the classical pathway in vitro. In contrast, complexes that have bound C3 in vivo do not contain IgM-RF and are weak complement activators in vitro [44]. JIA patients have been shown to have high levels of circulating immune complexes (CICs), which are correlated with disease activity. Low et al. concluded that JIA CICs might be used as a marker for increased B-cell activity [45]. Data from JIA patients suggested a scenario where different external antigens incite multiple antigen-specific pathways, cytotoxic T-cell responses, activation of the classic complement cascade, and production of pro-inflammatory cytokines [46]. Recent studies have demonstrated that complement activation and the overall levels of immune complexes are correlated with disease activity in JIA, thus indicating their role in the pathophysiology of the disease [47].

Immunological targets

Monoclonal antibodies are (largely) used in the treatment of neoplastic, autoimmune, and inflammatory diseases. Therefore, the clinical applications of monoclonal antibodies have become broader. Monoclonal antibody targets include, among others, CD20, HER-2, EGFR, IL-6 receptor, TNF-α, CD30, VEGF-A, IgE. Examples of immune-mediated and inflammatory diseases that respond to the monoclonal antibody treatment include RA, Crohn's disease, ulcerative colitis, JIA, psoriasis and psoriatic arthritis, Wegener's granulomatosis, microscopic polyangiitis, ankylosing spondylitis (AS), plaque psoriasis, and asthma [48]. Adalimumab, a human monoclonal antibody to tumour necrosis factor alpha (TNF-α), demonstrated efficacy and tolerability in patients suffering from many

inflammatory conditions such as RA, psoriatic arthritis, plaque psoriasis, inflammatory bowel diseases, ulcerative colitis, paediatric Crohn's disease, intestinal Behçet's disease, AS, axial spondyloarthritis, and JIA [49]. Adalimumab was initially approved (in 2002) for the treatment of moderate to severe RA. In the following years, its anti-inflammatory properties were applied to the pJIA treatment [50]. CT-P13 became the first monoclonal antibody biosimilar approved by the European Medicines Agency [51]. However, countless medical authorities disagree with the extrapolation of its prescription for JIA, due to its biosimilarity. It has only been tested in two disease models: AS and RA [52]. We have searched for recent/novel monoclonal Ab treatment for JIA in the 2017 literature and found the following: Braun-Moscovici et al. evaluated the serum infliximab (IFX) levels and levels of IFX-Ab in the management of rheumatic diseases. The most useful information for therapy was obtained in patients with low IFX levels and low levels of IFX-Ab [53]. Machado et al. evaluated the safety of TCZ in the treatment of JIA and found that TCZ was the best therapy for patients with severe forms of sJIA and pJIA, but further laboratory assessments of these patients were needed [54].

The enzymes in the Janus kinase (JAK) family are signalling molecules. JAK inhibitors are novel targets in sJIA [55]. To facitinib response, a JAK inhibitor, was studied in JIA [56] and was found to be safe in RA treatment, but it interferes with signal transduction via cytokine receptors using the common γ -chain [57], and clinical trials remain in an early phase [58].

Intravenous immune globulin (IVIG) infusions were effective in alleviating the systemic manifestations of sJIA but were less effective in controlling long-lasting arthritis for more than one year [59]. IVIG is a biologic immune-modulatory agent that operates through various mechanisms. Therefore, IVIG may be considered a potential tool for the treatment of juvenile chronic arthritis [60].

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References

- Park H, Bourla AB, Kastner DL, et al. (2012): Lighting the fires within: the cell biology of autoinflammatory diseases. Nat Rev Immunol 12: 570-580.
- Canna SW, Goldbach-Mansky R (2015): New monogenic autoinflammatory diseases – a clinical overview. Semin Immunopathol 37: 387-394.
- Ombrello MJ (2015): Advances in the genetically complex autoinflammatory diseases. Semin Immunopathol 37: 403-406
- 4. Hedrich CM (2016): Shaping the spectrum from autoinflammation to autoimmunity. Clin Immunol 165: 21-28.
- Hromadnikova I, Stechova K, Pavla V, et al. (2002): Anticyclic citrullinated peptide antibodies in patients with juvenile idiopathic arthritis. Autoimmunity 35: 397-401.

- Wu R, Shovman O, Zhang Y, et al. (2007): Increased prevalence of anti-third generation cyclic citrullinated peptide antibodies in patients with rheumatoid arthritis and CREST syndrome. Clin Rev Allergy Immunol 32: 47-56.
- Habib HM, Mosaad YM, Youssef HM (2008): Anti-cyclic citrullinated peptide antibodies in patients with juvenile idiopathic arthritis. Immunol Invest 37: 849-857.
- Zheng J, Yin J, Huang R, et al. (2013): Meta-analysis reveals an association of STAT4 polymorphisms with systemic autoimmune disorders and anti-dsDNA antibody. Hum Immunol 74: 986-992.
- Wang Y, Pei F, Wang X, et al. (2015): Meta-analysis: diagnostic accuracy of anti-cyclic citrullinated peptide antibody for juvenile idiopathic arthritis. J Immunol Res 2015: 915276.
- Nguyen H, James EA (2016): Immune recognition of citrullinated epitopes. Immunology 149: 131-138.
- 11. van der Zee R, Anderton SM, Prakken AB, et al. (1998): T cell responses to conserved bacterial heat-shock-protein epitopes induce resistance in experimental autoimmunity. Semin Immunol 10: 35-41.
- Zlacka D, Vavrincova P, Hien Nguyen TT, Hromadnikova I (2006): Frequency of anti-hsp60, -65 and -70 antibodies in sera of patients with juvenile idiopathic arthritis. J Autoimmun 27: 81-88.
- Nguyen TT, Bezouska K, Vavrincova P, et al. (2008): Humoral response against Mycobacterium bovis Hsp65 derived fragments in children and young people with various disorders.
 J Immunoassay Immunochem 29: 281-298.
- Durai M, Huang MN, Moudgil KD (2009): Self heat-shock protein 65-mediated regulation of autoimmune arthritis. J Autoimmun 33: 208-213.
- Lin YT, Wang CT, Gershwin ME, Chiang BL (2011): The pathogenesis of oligoarticular/polyarticular vs systemic juvenile idiopathic arthritis. Autoimmun Rev 10: 482-489.
- Kim EY, Durai M, Mia Y, et al. (2016): Modulation of adjuvant arthritis by cellular and humoral immunity to Hsp65. Front Immunol 7: 203.
- Bronson PG, Ramsay PP, Seldin MF, et al. (2010): A candidate gene study of CLEC16A does not provide evidence of association with risk for anti-CCP-positive rheumatoid arthritis. Genes Immun 11: 504-508.
- Schuster C, Gerold KD, Schober K, et al. (2015): The autoimmunity-associated gene CLEC16A modulates thymic epithelial cell autophagy and alters T cell selection. Immunity 42: 942-952.
- Yang CA, Chiang BL (2015): Inflammasomes and human autoimmunity: a comprehensive review. J Autoimmun 61: 1-8.
- Man SM, Karki R, Kanneganti TD (2016): AIM2 inflammasome in infection, cancer, and autoimmunity: Role in DNA sensing, inflammation, and innate immunity. Eur J Immunol 46: 269-280.
- Kessel C, Holzinger D, Foell D (2013): Phagocyte-derived S100 proteins in autoinflammation: putative role in pathogenesis and usefulness as biomarkers. Clin Immunol 147: 229-241.
- Bruck N, Schnabel A, Hedrich CM (2015): Current understanding of the pathophysiology of systemic juvenile idiopathic arthritis (sJIA) and target-directed therapeutic approaches. Clin Immunol 159: 72-83.
- Grom AA (2014): Canakinumab for the treatment of systemic juvenile idiopathic arthritis. Expert Rev Clin Immunol 10: 1427-1435.
- Davies R, Choy E (2014): Clinical experience of IL-6 blockade in rheumatic diseases – implications on IL-6 biology and disease pathogenesis. Semin Immunol 26: 97-104.

- 25. Kang S, Tanaka T, Kishimoto T (2015): Therapeutic uses of anti-interleukin-6 receptor antibody. Int Immunol 27: 21-29.
- Woerner A, von Scheven-Gete A, Cimaz R, Hofer M (2015): Complications of systemic juvenile idiopathic arthritis: risk factors and management recommendations. Expert Rev Clin Immunol 11: 575-588.
- Tanaka T, Narazaki M, Ogata A, Kishimoto T (2014): A new era for the treatment of inflammatory autoimmune diseases by interleukin-6 blockade strategy. Semin Immunol 26: 88-96.
- Barone P, Pignataro R, Garozzo MT, Leonardi S (2016): IL-6 blockers in systemic onset juvenile idiopathic arthritis. Immunotherapy 8: 79-87.
- 29. Harsini S, Ziaee V, Maddah M, et al. (2016): Interleukin 10 and transforming growth factor beta 1 gene polymorphisms in juvenile idiopathic arthritis. Bratisl Lek Listy 117: 258-262.
- 30. Fathy MM, Elsaadany HF, Ali YF, et al. (2017): Association of IL-10 gene polymorphisms and susceptibility to juvenile idiopathic arthritis in Egyptian children and adolescents: a case-control study. Ital J Pediatr 43: 9.
- Drozdova EA, Yadykina EV, Mezentseva EA, Nikushkina KV (2017): Cytokine profile changes in children with juvenile idiopathic arthritis-associated uveitis. Vestn Oftalmol 133: 27-31.
- 32. Kessel C, Lippitz K, Weinhage T, et al (2017): Proinflammatory cytokine environments can drive interleukin-17 overexpression by γ/δ T cells in systemic juvenile idiopathic arthritis. Arthritis Rheumatol 69: 1480-1494.
- 33. Inoue N, Shimizu M, Tsunoda S, et al. (2016): Cytokine profile in adult-onset Still's disease: comparison with systemic juvenile idiopathic arthritis. Clin Immunol 169: 8-13.
- 34. Swart JF, de Roock S, Prakken BJ (2016): Understanding inflammation in juvenile idiopathic arthritis: how immune biomarkers guide clinical strategies in the systemic onset subtype. Eur J Immunol 46: 2068-2077.
- 35. Shimizu M, Nakagishi Y, Inoue N, et al. (2015): Interleukin-18 for predicting the development of macrophage activation syndrome in systemic juvenile idiopathic arthritis. Clin Immunol 160: 277-281.
- Behrens EM (2008): Macrophage activation syndrome in rheumatic disease: what is the role of the antigen presenting cell? Autoimmun Rev 7: 305-308.
- 37. Ravelli A, Grom AA, Behrens EM, Cron RQ (2012): Macrophage activation syndrome as part of systemic juvenile idiopathic arthritis: diagnosis, genetics, pathophysiology and treatment. Genes Immun 13: 289-298.
- 38. de Matos CT, Berg L, Michaëlsson J, et al. (2007): Activating and inhibitory receptors on synovial fluid natural killer cells of arthritis patients: role of CD94/NKG2A in control of cytokine secretion. Immunology 122: 291-301.
- Avau A, Put K, Wouters CH, Matthys P (2015): Cytokine balance and cytokine-driven natural killer cell dysfunction in systemic juvenile idiopathic arthritis. Cytokine Growth Factor Rev 26: 35-45.
- 40. Gaur P, Misra R, Aggarwal A (2015): Natural killer cell and gamma delta T cell alterations in enthesitis related arthritis category of juvenile idiopathic arthritis. Clin Immunol 161: 163-169.
- 41. Popko K, Górska E (2015): The role of natural killer cells in pathogenesis of autoimmune diseases. Centr Eur J Immunol 40: 470-476.
- Sandberg G, Söder O, Stenvinkel C, et al. (1988): Thymocytotoxic antibodies in synovial fluid. J Clin Lab Immunol 25: 115-118.

- 43. Bonagura VR, Ilowite NT, Hatam L, et al. (1991): Expression of the major rheumatoid factor cross-reactive idiotype in pediatric patients with systemic lupus erythematosus. Clin Immunol Immunopathol 60: 232-243.
- 44. Jarvis JN, Diebold MM, Chadwell MK, et al. (1995): Composition and biological behaviour of immune complexes isolated from synovial fluid of patients with juvenile rheumatoid arthritis (JRA). Clin Exp Immunol 100: 514-518.
- 45. Low JM, Chauhan AK, Moore TL (2007): Abnormal kappa:lambda light chain ratio in circulating immune complexes as a marker for B cell activity in juvenile idiopathic arthritis. Scand J Immunol 65: 76-83.
- Rigante D, Bosco A, Esposito S (2015): The etiology of juvenile idiopathic arthritis. Clin Rev Allergy Immunol 49: 253-261.
- 47. Moore TL (2016): Immune complexes in juvenile idiopathic arthritis. Front Immunol 7: 177.
- 48. Bonamichi-Santos R, Castells M (2018): Diagnoses and management of drug hypersensitivity and anaphylaxis in cancer and chronic inflammatory diseases: reactions to taxanes and monoclonal antibodies. Clin Rev Allergy Immunol 54: 375-385
- Lapadula G, Marchesoni A, Armuzzi A, et al. (2014): Adalimumab in the treatment of immune-mediated diseases. Int J Immunopathol Pharmacol 27: 33-48.
- 50. Patel AS, Suarez LD, Rosh JR (2016): Adalimumab in pediatric Crohn's disease. Immunotherapy 8: 127-133.
- Müller-Ladner U, Hong S, Oh C, Taylor P (2015): Scientific rationale behind the development and approval of biosimilar infliximab (CT-P13) in Europe. Expert Rev Clin Immunol 11 Suppl 1: S5-14.
- 52. Azevedo VF, Meirelles Ede S, Kochen Jde A, et al. (2015): Recommendations on the use of biosimilars by the Brazilian Society of Rheumatology, Brazilian Society of Dermatology, Brazilian Federation of Gastroenterology and Brazilian Study Group on Inflammatory Bowel Disease – Focus on clinical evaluation of monoclonal antibodies and fusion proteins used in the treatment of autoimmune diseases. Autoimmun Rev 14: 769-773
- 53. Braun-Moscovici Y, Dagan A, Toledano K, et al. (2017): The input of measuring infliximab levels and levels of antibodies to infliximab in the management of patients with rheumatic diseases. Harefuah 156: 427-430 [Article in Hebrew].
- 54. Machado SH, Xavier RM (2017): Safety of tocilizumab in the treatment of juvenile idiopathic arthritis. Expert Opin Drug Saf 16: 493-500.
- Canny S, Mellins E (2017): New frontiers in the treatment of systemic juvenile idiopathic arthritis. F1000Res 6: 971.
- Tseng B, Amighi A, Bradford K, et al. (2016): Tofacitinib response in juvenile idiopathic arthritis (JIA) and collagenous colitis. J Clin Rheumatol 22: 446-448.
- 57. Rizzi M, Lorenzetti R, Fischer K, et al. (2017): Impact of tofacitinib treatment on human B-cells in vitro and in vivo. J Autoimmun 77: 55-66.
- Mauro A, Rigante D, Cimaz R (2017): Investigational drugs for treatment of juvenile idiopathic arthritis. Expert Opin Investig Drugs 26: 381-387.
- Roifman CM (1995): Use of intravenous immune globulin in the therapy of children with rheumatological diseases. J Clin Immunol 15 (6 Suppl): 42S-51S.
- Katz-Agranov N, Khattri S, Zandman-Goddard G (2015): The role of intravenous immunoglobulins in the treatment of rheumatoid arthritis. Autoimmun Rev 14: 651-658.